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Key indicators

Single-crystal X-ray study
T = 293 K
Mean $\sigma(\text{C}-\text{C}) = 0.004 \text{ \AA}$
R factor = 0.038
wR factor = 0.105
Data-to-parameter ratio = 8.2

For details of how these key indicators were automatically derived from the article, see <http://journals.iucr.org/e>.

A taxane diterpenoid from the needles of *Taxus wallichiana*

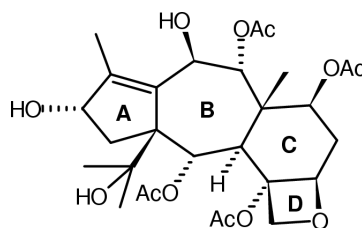
The title compound, 10,13-deacetyl-*abeo*-baccatin IV, $\text{C}_{28}\text{H}_{40}\text{O}_{12}$, crystallized in space group *P*1. The molecular structure shows that the *B/C* ring junction is *trans*-fused, while the *A* ring is in a *syn* conformation with respect to the *C* ring and *anti* with respect to the *D* ring. The conformations of the individual rings differ from each other. The molecule as a whole adopts a cage-type folded conformation. Intra- and intermolecular $\text{O}-\text{H}\cdots\text{O}$ and $\text{C}-\text{H}\cdots\text{O}$ hydrogen bonds, together with van der Waals interactions, stabilize the crystal structure.

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Comment

Investigations on the taxoid constituents of different parts of *Taxus wallichiana* led to isolation of the title compound, (I). Taxoid (I) was isolated from the needles of *T. wallichiana* (Chattopadhyay *et al.*, 1995). Considerable attention has been given to this type of diterpenoid molecule whose archetype is the paclitaxel (TaxolTM), a promising cancer chemotherapeutic agent (Appendino, 1995). Although numerous X-ray investigations have been performed on this and related molecules, there is much heated debate about the active conformation of this anticancer drug (Mastropaolo *et al.*, 1995, and references therein). This prompted us to undertake the present X-ray study of the title compound, (I), to determine its crystal structure and stereochemistry unequivocally.



(I)

Fig. 1 shows the structure of (I) with the atomic numbering scheme. Selected torsion angles of the terpenoid core of (I) are listed in Table 1. The molecule contains a three-ring fused system *A/B/C* with an additional *D* ring attached to the ring *C*. The *A* ring is in a *syn* conformation with respect to the *C* ring and *anti* with respect to the *D* ring. The *B/C* junction is *trans*-fused due to the *trans*-axial dispositions of C25 at C8 and H3 at C3. Thus, the molecule as a whole adopts a folded cage-type conformation. Least-squares plane calculations indicate that the seven-membered *B* ring adopts a boat conformation [the deviations of atoms C1, C2 and C9 are 1.078 (4), 1.289 (4) and 0.553 (4) Å, respectively from the least-squares plane through

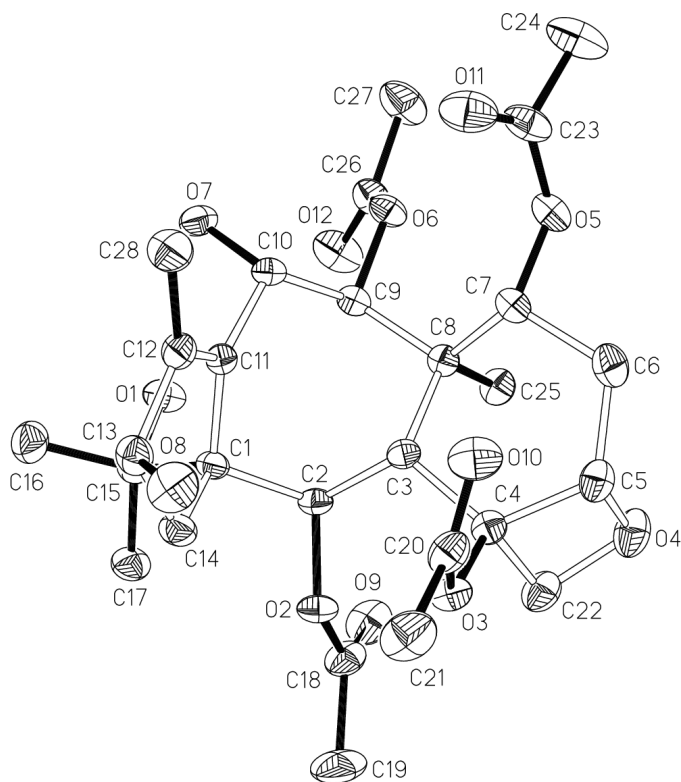


Figure 1

The molecular structure and crystallographic numbering scheme for (I). Displacement ellipsoids are shown at the 30% probability level. H atoms have been omitted for clarity.

atoms C3, C8, C10 and C11]. The six-membered C ring is in an envelope conformation while the five-membered A ring is puckered to form an envelope. The molecular packing in the crystal shows that the hydroxyl groups are involved in both intra- and intermolecular hydrogen bonding of the type O—H···O. In addition, the packing further reveals the presence of intra- and intermolecular C—H···O hydrogen bonding. Thus, O—H···O and C—H···O hydrogen-bonding interactions, along with van der Waals forces, stabilize the solid-state structure.

Experimental

Compound (I) was isolated from methanol extracts of the needles of *T. wallichiana* following the reported protocols (Chattopadhyay *et al.*, 1995). Diffraction quality crystals were grown at room temperature by slow evaporation of a methanolic solution.

Crystal data

$C_{28}H_{40}O_{12}$	$Z = 1$
$M_r = 568.60$	$D_x = 1.282 \text{ Mg m}^{-3}$
Triclinic, $P1$	Mo $K\alpha$ radiation
$a = 8.957 (1) \text{ \AA}$	Cell parameters from 26 reflections
$b = 9.667 (1) \text{ \AA}$	$\theta = 5.0\text{--}9.2^\circ$
$c = 9.827 (1) \text{ \AA}$	$\mu = 0.10 \text{ mm}^{-1}$
$\alpha = 110.10 (1)^\circ$	$T = 293 (2) \text{ K}$
$\beta = 92.42 (1)^\circ$	Block, colourless
$\gamma = 110.39 (1)^\circ$	$0.48 \times 0.35 \times 0.33 \text{ mm}$
$V = 736.2 (1) \text{ \AA}^3$	

Data collection

Bruker P4 diffractometer	$k = -10 \rightarrow 10$
θ - 2θ scans	$l = -11 \rightarrow 11$
3060 measured reflections	3 standard reflections
3060 independent reflections	every 97 reflections
2941 reflections with $I > 2\sigma(I)$	frequency: 60 min
$\theta_{\text{max}} = 25.0^\circ$	intensity decay: none
$h = -10 \rightarrow 1$	

Refinement

Refinement on F^2	$w = 1/[\sigma^2(F_o^2) + (0.0759P)^2 + 0.0885P]$
$R[F^2 > 2\sigma(F^2)] = 0.038$	where $P = (F_o^2 + 2F_c^2)/3$
$wR(F^2) = 0.105$	$(\Delta/\sigma)_{\text{max}} = 0.002$
$S = 1.04$	$\Delta\rho_{\text{max}} = 0.18 \text{ e \AA}^{-3}$
3060 reflections	$\Delta\rho_{\text{min}} = -0.18 \text{ e \AA}^{-3}$
372 parameters	
H-atom parameters constrained	

Table 1

Selected torsion angles ($^\circ$).

C11—C1—C2—C3	27.8 (3)	C3—C8—C9—C10	48.8 (3)
C14—C1—C2—C3	−82.9 (3)	C8—C9—C10—C11	−53.1 (3)
C15—C1—C2—C3	152.90 (19)	C9—C10—C11—C12	146.7 (2)
C1—C2—C3—C4	130.6 (2)	C9—C10—C11—C1	−33.9 (3)
C1—C2—C3—C8	−96.5 (2)	C14—C1—C11—C12	−11.9 (3)
C2—C3—C4—C22	58.0 (3)	C15—C1—C11—C12	106.5 (2)
C8—C3—C4—C22	−74.2 (3)	C2—C1—C11—C12	−128.7 (2)
C2—C3—C4—C5	159.8 (2)	C14—C1—C11—C10	168.6 (2)
C8—C3—C4—C5	27.5 (3)	C15—C1—C11—C10	−73.0 (3)
C22—C4—C5—O4	10.3 (2)	C2—C1—C11—C10	51.8 (3)
C3—C4—C5—O4	−112.2 (2)	C10—C11—C12—C13	−179.3 (2)
C22—C4—C5—C6	126.1 (3)	C1—C11—C12—C13	1.2 (3)
C3—C4—C5—C6	3.6 (4)	C11—C12—C13—C14	10.4 (3)
O4—C5—C6—C7	104.3 (3)	C12—C13—C14—C1	−17.7 (3)
C4—C5—C6—C7	1.0 (4)	C11—C1—C14—C13	17.9 (3)
C5—C6—C7—C8	−38.1 (3)	C15—C1—C14—C13	−100.9 (3)
C6—C7—C8—C9	−176.44 (19)	C2—C1—C14—C13	135.0 (2)
C6—C7—C8—C3	66.2 (2)	C5—C4—C22—O4	−10.4 (2)
C4—C3—C8—C7	−60.0 (3)	C3—C4—C22—O4	108.8 (3)
C2—C3—C8—C7	165.76 (19)	C4—C22—O4—C5	11.0 (2)
C4—C3—C8—C9	177.9 (2)	C6—C5—O4—C22	−132.9 (3)
C2—C3—C8—C9	43.7 (2)	C4—C5—O4—C22	−10.8 (2)
C7—C8—C9—C10	−66.2 (3)		

Table 2

Hydrogen-bonding geometry (\AA , $^\circ$).

$D\text{—}H\cdots A$	$D\text{—}H$	$H\cdots A$	$D\cdots A$	$D\text{—}H\cdots A$
O7—H7···O1	0.82	2.02	2.740 (3)	146
O1—H1···O11 ⁱ	0.82	2.08	2.883 (3)	168
O8—H8···O4 ⁱⁱ	0.82	2.08	2.845 (3)	154
O8—H8···O12 ⁱⁱⁱ	0.82	2.64	3.075 (3)	115
C6—H6B···O9 ^{iv}	0.97	2.42	3.366 (5)	166
C9—H9···O1	0.98	2.44	3.136 (3)	128
C25—H19C···O4	0.96	2.28	3.123 (4)	146
C22—H20B···O9	0.97	2.42	3.258 (5)	144
C24—H27C···O12 ^{iv}	0.96	2.48	3.443 (6)	175

Symmetry codes: (i) $1 + x, y, z$; (ii) $x, y, 1 + z$; (iii) $x, 1 + y, 1 + z$; (iv) $x - 1, y, z$.

Data collection: XSCANS (Siemens, 1996); cell refinement: XSCANS; data reduction: XSCANS; program(s) used to solve structure: SHELXTL-NT (Bruker, 1997); program(s) used to refine structure: SHELXTL-NT; molecular graphics: SHELXTL-NT; software used to prepare material for publication: SHELXTL-NT.

References

- Appendino, G. (1995). *Nat. Prod. Rep.* **12**, 349–360.
- Bruker (1997). *SHELXTL-NT*. Version 5.1. Bruker AXS Inc., Madison, Wisconsin, USA.
- Chattopadhyay, S. K., Sharma, R. P., Appendino, G. & Gariboldi, P. (1995). *Phytochemistry*, pp. 869–870.
- Mastropaolo, D., Camerman, A., Luo, Y., Brayer, G. D. & Camerman, N. (1995). *Proc. Natl Acad. Sci. USA*, **92**, 6920–6924.
- Siemens (1996). *XSCANS*. Version 2.21. Siemens Analytical X-ray Instruments Inc., Madison, Wisconsin, USA.